Quick Protocol

DNJia Virus DNA Kit

Viral DNA/RNA isolation based on silica technology

- MiniPrep

For Viral DNA Isolation from

Body Fluid
Serum
Plasma

Kit Content

Component	100 preps
GLB	20ml
BWB1 (concentrate)	2 x 16ml
BWB2 (concentrate)	2 x 12ml
ERR	5ml
RJ-Protease	2 x 1.25ml
HiPure DR Column	100
Collection Tube	200

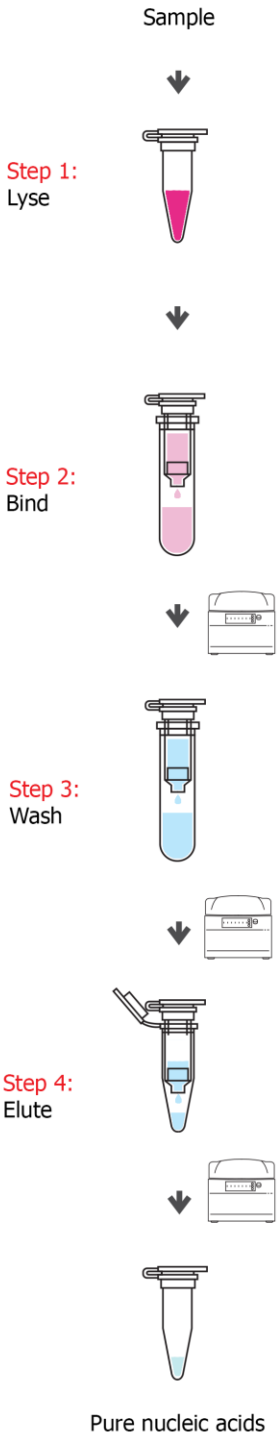
Washing Buffer Preparation

Before the first use, add appropriate amount of ethanol (96-100%) to each washing buffer tube, then mix thoroughly to prepare washing buffer, refer to Table 1. Do not forget to tick the check box on the bottle label to indicate that ethanol has been added. Before each use mix reconstituted buffer by shaking. Storing at room temperature.

Table 1: Washing buffer preparation

Buffer Name	Concentrated Volume	Amount of Ethanol	Final Volume
BWB1	16ml	24ml	40ml
BWB2	12ml	28ml	40ml

Procedure of silica-based DNA isolation in quick look



Protocol, Isolation of Viral DNA

Sample type:

- Body fluid
- Serum
- Plasma

Recommended Starting Material for Isolation of Viral DNA

The size of recommended starting material to use with determined lysis volume are listed here.

Table 2: Recommended starting material and Lysis Buffer amount

Sample	Size of Starting material	Lysis Buffer Amount
Plasma/serum	Up to 200µl	200µl

Some tips to know

- All centrifugation steps are carried out at room temperature (15–25°C).
- Do not forget to add the appropriate amount of molecular biology grade ethanol (96–100%) to BWB1 and BWB2 buffers as indicated on the bottle, before using for the first time, refer to washing buffer preparation.
- If GLB or TLB forms precipitate, please warm it to 56°C until the precipitate has fully dissolved.

Process

- Add 25µl RJ-Protease to a 1.5ml clean microcentrifuge tube. Add 200µl sample (plasma, serum, body fluid and etc.) to the tube. Then add 200µl GLB. Pulse vortex for 15s and incubate at 56 °C for 12min.
- Add 200µl ethanol (%96-100) to the lysate, mix by pulse vortexing for 15s, then centrifuge briefly.
- Gently, pipette the mixture to a HiPure DR column placed in a 2ml collection tube (supplied in the kit box). Centrifuge at 8000 rpm for 1min. Discard flow-through and place back the HiPure DR column in to the collection tube.
- Add 500µl BWB1 and centrifuge for 1min at 8000 rpm, discard both the flow-through and the collection tube. Place back the HiPure DR column in to the collection tube.

- Add 500µl BWB2 and centrifuge for 3min at 14000 rpm. Discard both the flow-through and the collection tube. Place the HiPure DR column in a new clean 1.5ml microcentrifuge tube (not provided).

  Note: To avoid ethanol carry over, be careful that the column does not come into contact with the flow-through, if it happens discard the flow-through, place the column back in a collection tube and centrifuge for another 1min at 14000 rpm.

- Pipette 30-50µl ERR directly onto HiPure DR column. Incubate at room temperature for 1-5min. Centrifuge it at 12000 rpm for 1min.