Quick Protocol

RNA Stabilization in Harvested Animal Tissues

process

- Cut the animal tissue sample into slices less than 5mm thick, as quickly as possible.
- Completely immerse the tissue pieces in the collection vessel containing RNaseLag.

  Note: Make sure to use the appropriate volume of RNaseLag, so weight your sample before starting the procedure and use 10µl RNaseLag per 1mg of tissue.
- The sample is ready for archival storage at conditions shown in Table 4.
- After storage, for RNA isolation continue with appropriate protocol for the chosen sample type.

Table 1: Storage conditions and procedures after RNaseLag treatment.

Storage condition	Protocol
2–8°C	Incubate the prepared sample (in RNaseLag) for up to 4 weeks at 2–8°C.
15-25°C	Incubate the prepared sample (in RNaseLag) for up to 7 days at 15-25°C.
37°C	Incubate the prepared sample (in RNaseLag) for up to 1 days at 37°C.
–20°C	First incubate the prepared sample (in the RNaseLag) overnight at 2–8°C. Then transfer it to –20°C for storage.
–80°C	First incubate the prepared sample (in the RNaseLag) at 2–8°C. Then remove the tissue from the reagent, and transfer it to –80°C for long storage.

Protocol RNA Stabilization in Harvested Bacterial Cells

Process

- Calculate the required volume of bacterial culture (refer to Appendix 4).
- Add 2 volumes of RNaseLag into a tube
- Add 1 volume of bacterial culture to the tube. Mix by vortexing for 5 sec. Incubate at room temperature (15–25°C) for 5 min.
- Centrifuge for 10 min at 4000 rpm at universal centrifuge.

  Note: Sometimes the pellet is too clear to be recognized, it is due to RNaseLag treatment, and will not affect the ongoing process.
- Decant the supernatant.

Pellets can be stored at –20 to –30 °C for up to 2 weeks or at –70 °C for up to one month. For RNA isolation, thaw pellets at room temperature (15–25 °C) and proceed the appropriate RNA isolation protocol.