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Recommended Starting Material

To reach optimized results it is better to follow as listed below. The size of the recommended material to use with determined RNSol and chloroform amount are written in Table 1.

Table 1. Appropriate sample size and amount of RNSol, Chloroform and Ethanol amount

| Sample Size | RNSol Amount | Chloroform Amount |
|---|--------------|-------------------|
| 10 mg animal tissue, 1 × 10 ⁶ animal cells | 100 µl | 100 µl |
| 20 mg animal tissue 2 × 10 ⁶ animal cells | 200 µl | 100 µl |
| 30 mg animal tissue 3 × 10 ⁶ animal cells | 300 µl | 200 µl |
| 40 mg animal tissue 4 × 10 ⁶ animal cells | 400 µl | 200 µl |
| 50 mg animal tissue 5 × 10 ⁶ animal cells 1-3 ml whole Blood | 500 µl | 300 µl |
| 60 mg animal tissue 6 × 10 ⁶ animal cells | 600 µl | 300 µl |
| 70 mg animal tissue 7 × 10 ⁶ animal cells 3-5 ml whole Blood | 700 µl | 400 µl |
| 80 mg animal tissue 8 × 10 ⁶ animal cells | 800 µl | 400 µl |
| 90 mg animal tissue 9 × 10 ⁶ animal cells | 900 µl | 500 µl |
| 100 mg animal tissue 10 ⁷ animal cells 5-10 ml whole Blood | 1ml | 500 µl |

Protocol 1

Isolation of Total RNA (based on silica technology)

Sample Type: Animal tissues (fresh and frozen)

Some tips to know:

- All steps, before applying sample into spin column, are carried out on ice.
- Not forget to add the appropriate amount of Ethanol (96–100%) to TW1 and TW2 buffers (TWB1 and TWB2) as indicated on the bottle, before using for the first time.
- For frozen samples, thaw them to room temperature. Avoid repeated thawing and freezing of samples due to decreasing in RNA yield.
- If working with RNA for the first time, read Appendix 1, in main handbook, carefully.

Process

1. Remove the tissue from RNaseLag or use fresh tissue. Determine the Weight of starting material and Add appropriate amount of RNSol Reagent to the tissue sample (refer to the Table 1).
2. Disrupt and homogenize the tissue sample by selecting one of these ways:
 - After adding appropriate amount of RNSol, use Micropestle followed by homogenizer or syringe needle to homogenize the tissue.
 - After adding appropriate amount of RNSol, using TissueLyser or homogenizer to disrupt and homogenize the sample simultaneously.
 - Grind the weighed tissue in liquid nitrogen carefully with a cold mortar and pestle. Transfer tissue powder into an RNase-free, liquid-nitrogen-cooled, 1.5 ml microcentrifuge tube. Let the liquid nitrogen to evaporate completely (but do

not allow the tissue to thaw). Add the appropriate volume of RNSol (refer to the Table 1) and homogenize by passing the lysate 5-10 times through a blunt 20-gauge needle fitted to an RNase-free syringe.

Optional: Homogenizing step is optional. It means that using homogenizer or syringe needle to homogenize the lysate can be omitted from the process.

3. Add appropriate amount of chloroform (refer to Table 1), vigorously shake it for 30 s. Then pulse vortex for 15 s and incubate at -20°C for 2 min.
4. Centrifuge at 4°C for 12 min at 13000 rpm.
5. Transfer the aqueous phase (the upper phase) to a new tube. Be careful to avoid interfering the interphase.
6. Add one and a half volume of Absolute Ethanol to separated aqueous phase. Pulse vortex for 30 s.
7. Transfer the solution to a spin column placed in a 2 ml collection tube (supplied in the kit box). Centrifuge for 1 min at 13000 rpm at room temperature. Discard the flow-through.
8. Add 700 μl TWB1 (TW1 buffer) to the spin column. Centrifuge for 1 min at 13000 rpm at room temperature. Discard the flow-through.
9. Add 500 μl TWB2 (TW2 buffer) to the spin column. Centrifuge for 1 min at 13000 rpm at room temperature. Discard the flow-through.
10. Add 500 μl TWB2 (TW2 buffer) to the spin column. Centrifuge for 3 min at 14000 rpm at room temperature. Discard the collection tube with the flow-through.

11. Place the spin column in a new 1.5 ml microtube. Add 30-100 μ l RNase-free water directly to the spin column membrane. Centrifuge for 1 min at 12000 rpm to elute the RNA.

Note: If the expected RNA yield is more than the yield from pervious step, put the spin column on a new microtube and add another 30-100 μ l RNase-free water. Centrifuge for 1 min at 12000 rpm. The yield will be nearly same as previous step. However, it is possible to pass the flow-through from step 11 once more to obtain RNA with higher concentration.

Protocol 2

Isolation of Total RNA (based on silica technology)

Sample Type: PBMC (Peripheral Blood Mononuclear Cell), Whole blood, WBC (White Blood Cell)

Some tips to know:

- All steps, before applying sample into spin column, are carried out on ice.
- Not forget to add the appropriate amount of Ethanol (96–100%) to TW1 and TW2 buffers (TWB1 and TWB2) as indicated on the bottle, before using for the first time.
- For frozen samples, thaw them to room temperature. Avoid repeated thawing and freezing of samples due to decreasing in RNA yield.
- If working with RNA for the first time, read Appendix 1, in main handbook, carefully.

Process

1. Collect 0.5 to 10 ml blood into EDTA tubes. Add 3 volume of RBC Lysis Buffer (RNase-free). Invert the tube 5 times and incubate at 4 °c for 10 min.
2. Pulse vortex every 2 min during incubation to intersperse the sample.
3. Collect the WBCs by centrifugation at 2700 x g for 10min at 4°C.
4. Discard the supernatant, add 2 volume of RBC Lysis Buffer (RNase-free) to the pellet, vortex until the pellet is dissolved completely.
5. Collect the WBCs by centrifugation at 2700 x g for 10 min at 4°C.
6. Discard the supernatant. Add appropriate amount of RNSol Reagent to the sample (refer to the table 1).

7. Disrupt and homogenize the sample by selecting one of these ways:

- After adding appropriate amount of RNSol, use Micropestle followed by homogenizer or syringe needle to homogenize the cell pellet.
- After adding appropriate amount of RNSol, use TissueLyser or homogenizer to disrupt and homogenize the sample simultaneously.

Optional: Homogenizing step is optional. It means that using homogenizer or syringe needle to homogenize the lysate can be omitted from the process.

8. After passing through the syringe for 5-10 times, pulse vortex for 1 min, incubate for 10 min at room temperature.

Note: During isolating RNA from PBMC, it is necessary to thoroughly homogenize the sample and it is recommended to homogenize by passing the lysate 5-10 times through a blunt 20-gauge needle fitted to an RNase-free syringe.

9. Add appropriate amount of chloroform (refer to Table 1), vigorously shake it for 30 s. Then pulse vortex for 15 s and incubate at room temperature for 5 min.

10. Centrifuge at 4°C for 12 min at 13000 rpm.

11. Transfer the aqueous phase (the upper phase) to a new tube. Be careful to avoid interfering the interphase.

12. Add 400 µl Absolute Ethanol to the separated aqueous phase (refer to Table 1). Pulse vortex for 30 s.

13. Transfer the solution to a spin column placed in a 2 ml collection tube (supplied in the kit box). Centrifuge for 1 min at 13000 rpm at room temperature. Discard the flow-through.
14. Add 700 μ l TWB1 (TW1 buffer) to the spin column. Centrifuge for 1 min at 13000 rpm at room temperature. Discard the flow-through.
15. Add 500 μ l TWB2 (TW2 buffer) to the spin column. Centrifuge for 1 min at 13000 rpm at room temperature. Discard the flow-through.
16. Add 500 μ l TWB2 (TW2 buffer) to the spin column. Centrifuge for 3 min at 14000 rpm at room temperature. Discard the collection tube with the flow-through.
17. Place the spin column in a new nuclease-free 1.5 ml microtube. Add 30-100 μ l RNase-free water directly to the spin column membrane. Centrifuge for 1 min at 12000 rpm to elute the RNA.

Note: If the expected RNA yield is more than the yield from previous step, put the spin column on a new microtube and add another 30-100 μ l RNase-free water. Centrifuge for 1 min at 12000 rpm. The yield will be nearly same as previous step. However, it is possible to pass the flow-through from step 17 once more to obtain RNA with higher concentration.

Protocol 3***Isolation of Total RNA (based on silica technology)*****Sample Type:** Animal cultured cell**Some tips to know:**

- All steps, before applying sample into spin column, are carried out on ice.
- Not forget to add the appropriate amount of Ethanol (96–100%) to TW1 and TW2 buffers (TWB1 and TWB2) as indicated on the bottle, before using for the first time.
- For frozen samples, thaw them to room temperature. Avoid repeated thawing and freezing of samples due to decreasing in RNA yield.
- If working with RNA for the first time, read Appendix 1, in main handbook, carefully.

Process

1. Remove the cultured cells from RNaseLag or use fresh cultured cells. Determine the number of cells. Pellet the appropriate number of cells by centrifuging for 5 min at 300 x g. Carefully remove all supernatant by aspiration (for information about sample preparation refer to table 2).

Table 2: Lysis of attached monolayer and suspension cells

| Cultured Cell Type | Lysis Type | Description |
|----------------------------|--|--|
| Cells grown in a monolayer | Lyse directly in the cell-culture vessel | Determine the number of cells. Completely aspirate the cell-culture medium. |
| | Trypsinize and collect cells at 300 x g | Determine the number of cells. Aspirate the medium, and wash the cells with PBS. Aspirate the PBS, and add 0.1–0.25% trypsin in PBS. After the cells detach from the |

| | | |
|---------------------------|-------------------------|--|
| | | surface, add medium (containing serum to inactivate the trypsin), transfer the cells to an RNase-free centrifuge tube, and centrifuge at 300 x g for 5 min. Completely aspirate the supernatant, and continue the process. |
| Cells grown in suspension | Collect cells at 300 xg | Determine the number of cells. Pellet the appropriate number of cells by centrifuging for 5 min at 300 x g. Carefully remove all supernatant by aspiration. |

Note: Incomplete removal of cell-culture medium will inhibit complete lysis and binding to spin column, which finally reduce RNA yield.

2. Disrupt the cells by selecting one of these ways:

a. Adding appropriate volume of RNSol (refer to Table 1), Vortex to mix.

Note: Before adding RNSol, flick the tube to loosen the cell pellet thoroughly. Make sure that loosening the cell occurs completely to avoid inefficient lysis and reduced RNA yields.

b. For direct lysis of cells grown in a monolayer, add the appropriate amount of RNSol to the cell-culture dish (refer to Table 1). Collect the lysate with a rubber policeman. Transfer the lysate into a microcentrifuge tube. Vortex to mix, and ensure that no cell clumps are visible.

3. Homogenize the lysate by:

a. Homogenize the lysate for 30 s using a homogenizer.

b. Pass the lysate at least 5 times through a blunt 20-gauge needle fitted to an RNase-free syringe.

c. Pass the lysate through a shredder spin column by centrifuging at full speed for 2 min.

Optional: Homogenizing step is optional. It means that using homogenizer or syringe needle to homogenize the lysate can be omitted from the process.

4. Add appropriate amount of chloroform (refer to Table 1), vigorously shake it for 30

s. Then pulse vortex for 15 s and incubate at -20°C for 2 min.

5. Centrifuge at 4°C for 12 min at 13000 rpm.

6. Transfer the aqueous phase (the upper phase) to a new tube. Be careful to avoid interfering the interphase.
7. Add one and a half volume of Absolute Ethanol to the separated aqueous phase. Pulse vortex for 30 s.
8. Transfer the solution to a spin column placed in a 2 ml collection tube (supplied in the kit box). Centrifuge for 1 min at 13000 rpm at room temperature. Discard the flow-through.
9. Add 700 μ l TWB1 (TW1 buffer) to the spin column. Centrifuge for 1 min at 13000 rpm at room temperature. Discard the flow-through.
10. Add 500 μ l TWB2 (TW2 buffer) to the spin column. Centrifuge for 1 min at 13000 rpm at room temperature. Discard the flow-through.
11. Add 500 μ l TWB2 (TW2 buffer) to the spin column. Centrifuge for 3 min at 14000 rpm at room temperature. Discard the collection tube with the flow-through.
12. Place the spin column in a new 1.5 ml microtube. Add 30-100 μ l RNase-free water directly to the spin column membrane. Centrifuge for 1 min at 12000 rpm to elute the RNA.

Note: If the expected RNA yield is more than the yield from previous step, put the spin column on a new microtube and add another 30-100 μ l RNase-free water. Centrifuge for 1 min at 12000 rpm. The yield will be nearly same as previous step. However, it is possible to pass the flow-through from step 12 once more to obtain RNA with higher concentration.